

Original Article

LOCALIZATION OF NERVE GROWTH FACTOR DURING POSTNATAL DEVELOPMENT OF THE RAT RETINA

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Summary

Nerve growth factor (NGF) is known to stimulate neurite outgrowth and support neuronal survival during retinal development and regeneration through the high-affinity tyrosine kinase (Trk) receptors. On the other hand, NGF binding to the low-affinity neurotrophin receptor p75 might induce programmed death in early phase of retinal development. We have studied the expression of the nerve growth factor in the postnatal development rat retina. We investigated NGF in retinas of rats on postnatal day 1, 3, 5, 7, 9, 11, 16, 30. Changes in expression of NGF and localization in developing retina were assessed by immunohistochemistry method. During early retina's development NGF is localized in ganglion cells, Muller and pigment cells and its expression increased during next days. Amacrine, horizontal and bipolar cells express NGF lately. The expression of NGF can be demonstrated in inner segments and external limiting membrane about day 5. The distribution of NGF in postnatal rat retina suggests that it plays role in neurite outgrowth, regulation of number of ganglion cells and synaptogenesis in inner retina. NGF take part in differentiation of inner segments of photoreceptor cells.

Key words: NGF, neurotrophins, retina, postnatal

Introduction

NGF was the first neurotrophic agent to be discovered and has become paradigm of factors preventing cell death in selective populations of neurons during the development of central and peripheral nervous system. The trophic signals elicited by NGF are transduced through its specific tyrosine kinase receptor TrkA and modulated by the common neurotrophin receptor p75 [1].

We investigated the expression of NGF in the postnatal development rat retina.

Materials and Methods

Animals and tissue preparation

Wistar rats of postnatal days 1(P1), 3(P3), 5(P5), 7(P7), 9(P9), 11(P11), 16 (P16) and 30(P30) were obtained from a commercial source and after anesthesia with ether were decapitated. Eyes were enucleated, briefly fixed in Carnoa, and after removal of the anterior segment the eyes were returned to fixation for 2 hr. According to the routine method the fixated eyes were embedded in paraffin and 4-5 µm sections were cut on microtome.

Immunohistochemistry

Deparaffined sections were incubated overnight at 4°C with a 1:100 dilution rabbit polyclonal antirabbit NGF

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antibody. The ABC kit (Santa Cruz Biotechnology, California) was used to visualize the NGF antibody by using the peroxidase-3-3'-diaminobenzidine (DAB) reaction according to the manufacturer's instructions.

Results

The differentiation and definite localization of rat retinal cell populations became during the first four weeks after birth. At P1 was clearly seen the multiple ganglion layer (RGL), displayed newly formed inner plexiform layer (IPL) and fine positive cells in the ventricular layer and processes of pigmental cells (PC) and blood vessels in nerve fibers (NL) (Fig. 1).

The inner segments of photoreceptors, body's cones and newly forming outer plexiform layer (OPL) showed the positive reaction at P5 (Fig. 2).

The immunoreactivity for NGF was increased in cell membranes of INL and ONL, inner segments of photoreceptors and outer limited membrane (MLE) at P9 (Fig. 3).

At P11 NGF expressed in blood vessels in INL and well painting was visualized in horizontal cells (HC), bipolar cells (BC), inner segments (IS) and outer limited membrane MLE (Fig. 4).

At P16 and P30 strong reaction was seen in retinal ganglion cells (RGC), PC, IPL, some AC, MLE and IS

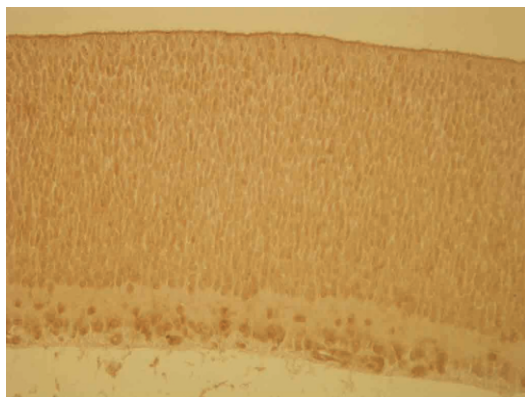


Figure 1. Expression of NGF in P1 rat retina. Labeling for NGF is most prominent in GCL. Labeling is also detected in IPL, ventricular layer and pigmental cells (Magnification x 400).

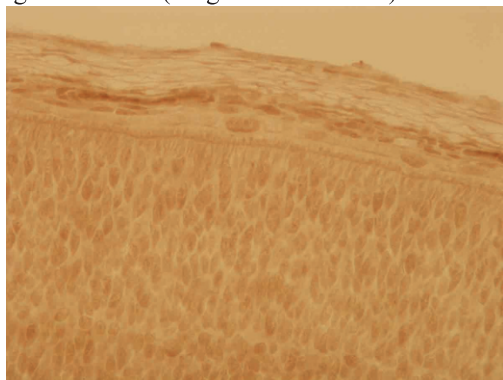


Figure 2. Expression of NGF in P5 rat retina. Inner segments of photoreceptors, body's cones and OPL are positive (Magnification x 400).

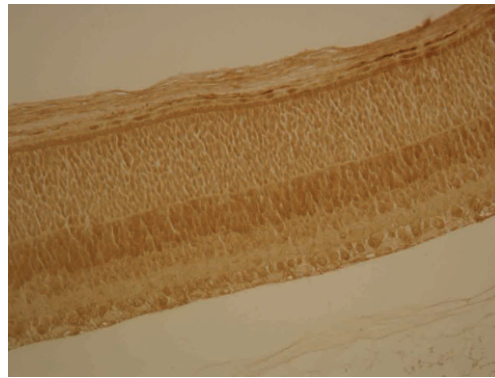


Figure 3. Expression of NGF in P9 rat retina. NGF immunoreactivity increases in plexiform layers and inner segments (Magnification x 400).

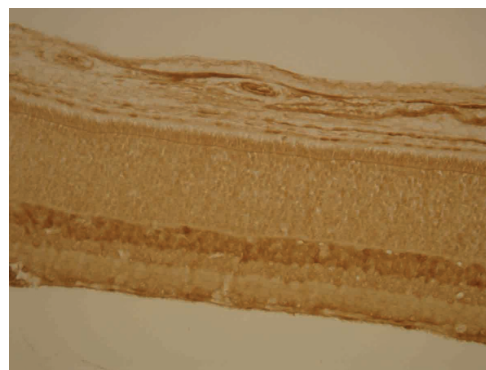


Figure 4. Expression of NGF in P11 rat retina. The immunoreactivity in body's of bipolar, horizontal cells can be seen in the external part of INL (Magnification x 400).

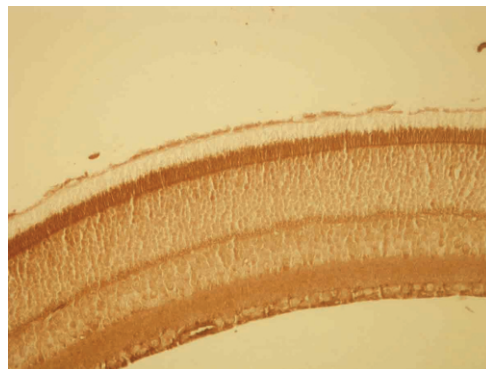


Figure 5. Expression of NGF in P30 rat retina. NGF immunoreactivity in RGL, IPL, OPL and IS are well visible (Magnification x 400).

Discussion

Programmed cell death occurs naturally, as a physiological process, during the embryonic development of multicellular organism. In retina, which belongs to the central nervous system, at least two phases of cell death have been reported to occur during development. An early phase takes place concomitant with the processes of neurogenesis, cell migration and cell differentiation. A later phase affecting mainly neurons occurs when connections are established and synapses

are formed, resulting in selective elimination of inappropriate connections. NGF controls the programmed cell death through binding to the p75 receptor which affects early postmitotic neuroblasts during development. On the other hand, retinal cells may survive the apoptotic effect of NGF by expression of the high-affinity TrkA receptor, which switches the pro-apoptotic signaling of P75 into a neurotrophic one [2, 3, 4].

The differentiation of rat retina is considered with physiological death of retina cells in postnatal development. The normal RGC death period in rat extends from around birth to 6 days later and is especially frequent between postnatal days 1 to 4. The presence of degenerating bipolar cells, amacrine cells and rod has been described at P4 in rat retinas. AC death does not cease until P26 and BC degeneration continues for 48 days. With respect to photoreceptors cells death, a second phase of apoptosis was reported between days 12 and 72, peaking at P23 [5, 6, 7].

We observed the expression of NGF in layered RGC on P1, which increased in the next days. The GCL was composed of layers at P0 and P5, but its thickness decreased and it becomes a monolayer by P17. This suggested a role of NGF in processes of regulation of number of RGC. The stratified RGC in chick retina themselves control their number by secreting NGF, which kills the incoming migratory RGC via p75 on their surface. RGC number control is different between mouse and chick retina. P75 and TrkA seem to be involved in regulation of mouse RGC number in early phase of retinal development, but they may be later adjusted by other molecules [8, 4].

The early positive reaction was found in ventricular layer, from which the photoreceptors (FC), horizontal (HC), bipolar (BC) and amacrine (AC) cells begin their differentiation. In mature rat retina the external parts of INL were positive and the body of BC cells was well visible. Karisson M et al [9, 10] found that NGF plays a role in horizontal cell plasticity and in the survival of horizontal cells (in autocrine mode of action) in avian retina but doesn't take part in survival of AC. We observed the decreasing of expression of AC and well positive BC in rat retina. NGF takes part in differentiation of BC in rat retina.

Very interesting fact was the early expression in the inner segment of photoreceptors, accordingly their differentiation and formation. NGF modulates secondary trophic factor expression in Müller cells that may contribute to protection of photoreceptors or increase their apoptosis [11].

Our investigation of expression in IPL and OPL suggested the role of NGF in synaptic plasticity and neurite outgrowth. It would be beneficial this NGF in the rat retina to go under some further investigations because it takes part in degenerating processes [11, 12] but intravitreal injection of endogenous NGF can protect retinal cells from degeneration in experimental retinal detachment [13].

Conclusion

The distribution of NGF in postnatal rat retina

suggests that it plays role in neurite outgrowth, regulation of number of ganglion cells and synaptogenesis in inner retina. NGF takes part in differentiation of inner segments of photoreceptor cells.

References

1. Lewin GR, Barde YA. Physiology of the neurotrophins. *Rev Neurosci*. 1996;19:289-317.
2. Dechnt G, Barde YA. Signaling through the neurotrophin receptor p75NTR. *Curr Opin Neurobiol*. 1997;7:413-418.
3. Frade JM. Unscheduled re-entry into the cell cycle induced by NGF precedes cell death in nascent retinal neurons. *J Cell Sci*. 2000;113:1139-1148.
4. Gonzales-Hoyuela M, Barbas A, Rodriguez-Tebar A. The autoregulation of retinal ganglion cell number. *Development*. 2000;128:117-124.
5. Vogel M, Muller K. Cellular decay in the rat retina during normal post-natal development: A preliminary quantitative analysis of the basic endogenous rhythm. *Exp Ophthalmol*. 1980;212:243-260.
6. Vouchidolova. Yochkova 1996. Постнатално развитие на клетъчните типове във вътрешните слоеве на ретината на плъх. *Българска Медицина*. 1996;5-6:63-66.
7. Vecino E, Hernandez M, Garsia M. Cell death in developing vertebrate retina. *Int J Dev Biol*. 2004;48:965-974.
8. Harada C, Harada T, Nakamya K, sakai Y, tanaka K, Parada L. Effect of p75NTR on the regulation of naturally occurring cell death and retinal ganglion number in the mouse eye. *Dev Biol*. 2006; 290(1): 57-65.
9. Karlsson Miriam, Mayodoro R, Reichardt LF, Catsicas S, Karten HJ, et al. Nerve Growth Factor is expressed by postmitotic avian retinal horizontal cells and support their survival during development in autocrine mode of action. *Development*. 2001;128,471-479.
10. Karlsson Miriam, Clary DO, Lefcort FB, Reichardt LF, Karten HJ, et al. Nerve Growth Factor Receptor TrkA Is Expressed by Horizontal Cells During the Chicken Retinal Development. *J Comp Neurol*. 1998;400:408-416.
11. Harada T, Harada C, Kohsaka S, Wada E, Yoshida K, et al. Microglia-Müller Glia cell Interactions Control Neurotrophic Factor Production During Light-Induced Retinal Degeneration. *J Neurosci*. 2002;22(21):9228-9236.
12. Ali TK, Matragon S, Pilla B, Liou G, El-Resemssy A, peroxynitrite Mediates Retinal Neurodegeneration by Inhibiting Nerve Growth Factor Survival Signaling in Experimental Human Diabetes. *Diabetes*. 2008;57(4):889-898.
13. Sun X, X Xu, F Wang, X Zhang, P Ho et al. Nerve Growth Factor Help Protect Retina in Experimental Retinal Detachment. *Ophthalmologica*. 2008;222:58-61.