

SELENIUM DEFICIENCY AS A RISK FACTOR FOR DEVELOPMENT OF ANEMIA

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Summary

Anemia is an important public health problem worldwide. Although iron (Fe) deficiency is considered the main factor in the pathogenesis of anemia, only 40-60% of anemia cases are responsive to Fe supplementation. Considerable data exist that other micronutrient deficiencies, such as selenium (Se), could be possible causes of anemia. The issue of Se deficiency as a risk factor for the development of anemia is of particular interest to our country since the Balkan region is known by a low Se content of soils. The aim of the study was to examine the contemporary conception of the influence of Se deficiency on the development of anemia by a review of the scientific literature. Most animal studies have shown a significant relation between Se deficiency and anemia, but one study indicates that there is no impact of Se deficiency on the hematological parameters. Associations of low serum Se with anemia have been found in a number of human studies including subjects of various age groups and pathological conditions. Three possible biological mechanisms have been suggested for the involvement of Se deficiency in the development of anemia: increased oxidative stress, modulation of inflammation through induction of interleukin-6, and increased expression of heme oxygenase-1. A more categorical clarification of the relationships between Se deficiency and development of anemia is needed with respect to appropriate trace element supplementation in cases of anemia with insufficient or absent therapeutic response to Fe treatment.

Key words: selenium deficiency, iron, anemia

Introduction

Anemia still remains an important public health problem worldwide. Iron deficiency anemia (IDA) is one of the ten major risk factors contributing to the global burden of diseases [1]. The most affected by anemia groups are children under 5 years of age (global prevalence of anemia 42.6%) and women of reproductive age (overall prevalence of anemia 29.4%), especially during pregnancy (38.2%) [2].

Although deficiency of iron (Fe) is considered the main factor in the pathogenesis of anemia, only 40-60% of anemia cases are responsive to treatment with Fe-containing medications and a large proportion of anemia

does not respond to iron supplementation [3]. Considerable data exist that other micronutrient deficiencies, such as copper (Cu) and selenium (Se), could be possible causes of anemia. The mechanisms by which Se deficiency is implicated in the pathogenesis of anemia have not been well characterized. The issue of Se deficiency as a risk factor for development of anemia is of particular interest to our country since the Balkan region is known by a low Se content of soils [4]. Poor soil Se levels determine lower dietary intakes of Se through food and drinking water.

The alarming fact is that the mean serum Se concentration for Bulgarian population (45 ± 6 $\mu\text{g/L}$) is lower compared to the Western European populations (70-120 $\mu\text{g/L}$) [5]. Results obtained by Lozanov et al. (2008) and Bivolarska (2014) have demonstrated the presence of Se deficiency in pregnant women living in some endemic regions of South Bulgaria - towns of Smolyan, Devin, Plovdiv, and Asenovgrad [6, 7]. A moderate Se deficiency has been found in school-age children from 7 to 10 years living in Smolyan endemic district [8].

Selenium is an essential trace element in all known forms of life. It is a cofactor of selenoproteins. The total number of known human selenoproteins is over 25, most of them with enzymatic activities. Selenoproteins are classified into 3 families: glutathione peroxidases (GPx), thioredoxin reductases (TrxR), and iodothyronine deiodinases (DIO). Selenoproteins participate in the defense against oxidative stress, muscle development and function, synthesis and metabolism of thyroid hormones. Selenoproteins enhance the apoptosis of cancer cells, improve the immune response against infectious diseases, suppress prostaglandin synthesis, and are involved in the normal sperm maturation and motility [9].

Selenoprotein expression is predominantly controlled by an adequate Se intake [10]. Selenium enters the body mainly by food [10, 11]. The major food sources of Se are Brazil nuts, organ meats, fish, poultry, and eggs. Good plant sources of Se are grains and vegetables grown on rich in Se soils, and sunflower seeds. It is worth noting that a diet containing 0.1 $\mu\text{g Se/g}$ food is considered sufficient to support normal growth and reproductive performance in all mammalian species [11].

The most widely used biomarker for assessment of Se status is the plasma or serum concentration of Se [10]. In 1997, Rayman suggested a value of 100 ng Se/mL serum as a criterion of adequate Se intake [12]. Serum Se values above 100 $\mu\text{g/L}$ are thought to be required for the optimal activity of GPx-1, a ubiquitously expressed cytosolic enzyme. Daniels (2004) has pointed out plasma Se concentration of 70-100 $\mu\text{g/L}$ as a threshold level beyond which further Se supplementation will produce no detectable increase in plasma GPx activity [10].

As a cofactor of selenoproteins, Se acts in antioxidant and redox processes of the organism [9, 10], including erythropoiesis and erythrocyte functioning [13-15]. Mechanisms by which Se influences Fe metabolism, synthesis of hemoglobin (Hb) and erythropoiesis have not yet been completely elucidated.

Our aim is to examine the contemporary conception of the influence of Se deficiency on the development of anemia by a review of the scientific literature.

Materials and Methods

A review of the world scientific literature was conducted. Data were gathered through the databases Google Scholar and PubMed for the time period from January 1980 to April 2017 by using the keywords: "selenium deficiency", "iron", and "anemia".

Experimental studies on the relationship between selenium deficiency and development of anemia in animals

The role of Se deficiency as a factor contributing to the development of anemia has been extensively studied in animal subjects. Several research groups have found that there is a significant relation between dietary Se deficiency and development of anemia in mice, rats and cattle [13, 15-17]. Anemia has been associated with enhanced susceptibility of erythrocytes to hemolysis and the presence of Heinz bodies, which is indicative of increased oxidation of Hb to methemoglobin (metHb) and denaturation of its globin subunits. Se supplementation has been shown to prevent hemolysis and formation

of Heinz bodies, elevate the values of Hb concentration and hematocrit (Hct), and correct the anemia. As opposed to these findings, Hu et al. (1984) have not observed any impact of the long-term Se deficiency on the hematological parameters in rats. No alterations have been found regarding the erythrocyte count, Hb concentration, Hct, and the osmotic resistance of erythrocytes. Likewise, results of the same study indicate that Se supplementation produces no significant effect on the examined hematological parameters [18].

Studies on the relationship between low serum levels of selenium and development of anemia in humans

Associations of low serum Se concentrations with anemia have been found in a number of human studies including subjects of various age groups: children of primary school age from 6 to 9 years [3, 19]; adolescent girls from 11 to 17 years of age [20]; adults aged 65 years and over [21-23]. Statistically significant relationships between low serum Se and development of anemia have also been observed in different pathological conditions, such as chronic kidney disease [24] and pulmonary tuberculosis [25]. Contrary to these results, studies in patients with IDA and control subjects have shown no relation between the serum Se level and Hb concentration [26, 27]. Comparative analysis of results between the studies investigating the relationship of low serum Se concentrations with anemia is presented in Table 1.

Interactions between selenium and iron

In order to gain insight into the mechanisms by which Se deficiency leads to development of anemia, particular attention should be paid on the interactions between Se and Fe. Results of studies on the relationships between serum Se concentrations and laboratory parameters characterizing Fe metabolism are inconsistent. Significantly lower serum levels of Se have been described in subjects with IDA compared to controls [26-29]. On the contrary, in young women with prelatent iron deficiency without

manifest anemia, serum concentrations of Se have not been found to differ significantly from those of controls [30]. No relationship has been observed between serum concentrations of Se and parameters assessing different aspects of iron metabolism, such as serum levels of ferritin, Fe, and TIBC, in both subjects with normal Fe status and patients with IDA [26, 27, 31].

Of particular interest are findings from interventional studies investigating the interactions between these two essential trace elements. Results of studies examining the effects of Se supplementation on body Fe status are contradictory. Morris et al. (1984) have reported a positive influence of Se supplementation on Hb concentration and Hct in cattle [16]. Conversely, studies by other research groups have not shown significant impact of Se supplementation on the values of Hb concentration, Hct, and serum ferritin [18, 31].

Interventional studies assessing the effects of Fe supplementation on body Se status also provide conflicting results. Some authors have reported for negative effects of supplementary Fe on the status of Se reflected by plasma Se concentration and serum GPx activity [31, 32]. Findings by other authors indicate that Fe supplementation has not impact on serum Se and GPx concentrations [30].

Influence of selenium on synthesis of hemoglobin and erythropoiesis

Findings from a number of studies indicate that Se stimulates synthesis of Hb and is essential for the processes of erythropoiesis. Oster et al. (1988) have found positive correlations of the cellular Se content of erythrocytes with the erythrocyte count, Hb concentration, and Hct [33]. Consistent with these results, Se supplementation has been found to induce a striking reticulocyte response [33] and to elevate the values of Hb concentration and Hct [16]. The significance of Se for the erythropoiesis is supported by the high levels of GPx present in plasma and erythrocytes [15].

Current understanding for the essential role of selenoproteins in erythropoiesis is provided by the studies of Kaushal et al. These authors have demonstrated that adequate cellular Se status

Table 1. Comparative analysis of results between studies investigating the relationship of low serum Se concentrations with anemia

| Author /Year | Design | Setting | Health status | Number of patients (n) | Demographic characteristics Age (years)* Sex Female (%) | Anemia (%)† | Serum/plasma Se (µmol/L) | Se deficiency (%)‡ | Relationship of Se deficiency with anemia §§ |
|-------------------------------|-----------------|---|---|--|--|-------------|---|--------------------|--|
| Van Nhien et al. (2008) [3] | Cross-sectional | Bac Ninh province, Vietnam | free from chronic and acute illness and congenital abnormality | 292 | 7.78±0.8 (range 6-9) 52 | 45.2 | 0.773±0.172 | 75.6 | p<0.05 |
| Rashid & Ghaznavi (2015) [19] | Cross-sectional | Nishtar Town Lahore, Pakistan | free from any acute or chronic illness | 150 | 7.56±1.2 (range 6-9) 45.4 | 36.7 | 0.489±0.022 | 48 | p<0.01 |
| Van Nhien et al. (2009) [20] | Cross-sectional | Duy Tien district, Ha Nam province, Vietnam | free from chronic and acute illnesses and congenital abnormalities | 245 | 14.1±2.1 (range 11-17) 100 | 20.4 | 1.36±0.432 | 15.9 | p<0.0001 |
| Bates et al. (2002) [21] | Cross-sectional | mainland Britain 1. subjects living in the community 2. subjects living in institutions | NA†† | total 1134 free-living 883 living in institutions 251 | ≥65 NA NA | NA | 0.90 (0.50-1.36)‡‡ significantly higher in free-living people than in those living in institutions | NA | p<0.05 |
| Semba et al. (2006) [22] | Cross-sectional | Baltimore (MD), USA subjects living in the community | stratified by group: 1. non-anemic 2. anemic significantly higher proportion of subjects from gr. 2 had diabetes mellitus, cardiovascular disease, positive cytomegalovirus serostatus, and elevated serum IL-6 compared to gr. 1 (p<0.05) | total 632 gr. 1 543 gr. 2 89 | range 70-79 mean=74.1 mean=74.6 | 14.1 | 1.55 (1.52-1.57)‡‡ 1.42 (1.37-1.47) p<0.0001 | NA | p=0.002 |
| Semba et al. (2009) [23] | Cross-sectional | USA non-institutionalized adults | stratified by group: 1. non-anemic 2. anemic significantly higher proportion of subjects from gr. 2 had osteoarthritis, diabetes and renal disease compared to gr. 1 (p<0.05) | total 2092 gr. 1 1822 gr. 2 270 | ≥65 73.6±0.2 75.2±0.6 | 12.9 | 1.60 (1.59-1.61)‡‡ 1.51 (1.47-1.56) p=0.0003 | NA | p=0.03 |

| Hampel et al. (1985) [24] | Case-control | NA | dialysis patients healthy controls | NA | NA | NA | NA | NA | NA | significantly lower in dialysis patients than in controls | NA | p<0.05 for dialysis group |
|-------------------------------|-----------------|------------------------|--|--|---|---|--------------------------------------|----|----|---|----|--|
| Van Lettow et al. (2005) [25] | Cross-sectional | Zomba district, Malawi | purposive sampling of adults with pulmonary tuberculosis stratified by group: 1. HIV-negative a) no anemia §§ b) mild to moderate anemia††† c) moderate to severe anemia†††† | total gr. 1a gr. 1b gr. 1c gr. 2 gr. 2a gr. 2b gr. 2c | 500 130 30 81 19 370 43 216 111 | mean=32 32±10 32±12 30±11 mean=34 32±8 34±9 34±8 | 54.6 76.9 33.3 49.4 47.4 | NA | NA | 0.729±0.23 0.639±0.25 0.554±0.22 p=0.01 0.724±0.22 0.633±0.20 0.513±0.20 p=0.001 | NA | p=0.001 for moderate to severe anemia p=0.006 |
| Yeğin et al. (1992) [26] | Case-control | Turkey | patients with IDA control subjects | total IDA gr. controls | 80 40 40 | children | NA | 50 | NA | significantly lower in the patient group (p<0.001) | NA | p>0.05 |
| Gürgöze et al. (2004) [27] | Case-control | Turkey | patients with IDA control subjects | total IDA gr. controls | 104 56 48 | range 1-8 | NA | 54 | NA | 0.426±0.104 0.709±0.215 p<0.001 | NA | p>0.05 |

*Values are presented as mean ± standard deviation unless otherwise indicated

†Defined as Hb concentrations below recommended thresholds for the respective sex and age group

‡Defined as serum Se concentration <0.89 µmol/L

§ Statistical significance is indicated by p<0.05

††NA – not available

‡‡Presented as mean (95% confidence interval)

§ § Hb concentration ≥120 g/L for females and ≥130 g/L for males;

†††Hb concentration <120 g/L for females and <130 g/L for males, and ≥80 g/L for both sexes;

‡‡‡Hb concentration <80 g/L for both sexes

regulates oxidative stress-dependent expression and activation of two transcription factors in erythroid cells, hypoxia-inducible factor (HIF)-1 α and Forkhead box O3 transcription factor (FoxO3a) [15, 34]. FoxO3a is known to be involved in the physiologic response to oxidative stress during erythropoiesis via induction of gene expression of antioxidant enzymes, including GPx-1, in erythroid precursors [35].

There is a significant body of research indicating that Se regulates biosynthesis and metabolism of heme [33, 36-38]. It has been found that Se induces expression of both the mitochondrial enzyme synthase of δ -aminolevulinic acid (δ -ALA) and the microsomal enzyme heme oxygenase-1. The latter is believed to be an indirect effect resulting from the increased production of “free” heme in the cell. Other enzymes involved in the biosynthesis of prosthetic group of Hb, such as dehydratase of δ -ALA and ferrochelatase, have shown to be significantly inhibited by Se.

Mechanisms by which Se deficiency can contribute to the development of anemia

Three possible biological mechanisms have been suggested for the involvement of Se deficiency in the development of anemia: increased oxidative stress, modulation of inflammation through induction of the proinflammatory cytokine interleukin (IL)-6, and increased expression of heme oxygenase-1 [3, 19].

Se-deficiency-induced oxidative stress

Data from experimental studies [13, 15-17] have shown that anemia in Se deficiency is associated with increased lipid peroxidation, elevated levels of protein carbonyls and metHb in the erythrocytes, denaturation of Hb, increased number of Heinz bodies, and decreased osmotic resistance of erythrocytes. These findings suggest increased generation of reactive oxygen species and exposure of erythrocytes to high degree of oxidative stress.

Selenium is a cofactor of GPx catalyzing the reduction of hydrogen peroxide and different organic peroxides [9, 14]. Activity of cytosolic GPx-1, which is responsible for the protection

of Hb in erythrocytes from oxidative stress, is diminished in Se deficiency [13, 15, 17, 18]. Increased lipid peroxidation of membrane lipids and denaturation of Hb can both contribute to hemolysis and reduced life-span of circulating erythrocytes. Moreover, recent study has shown that Se deficiency-induced anemia is associated with expansion of erythroid progenitors, but is not accompanied by an increased reticulocyte count. These results indicate inefficient erythropoiesis with defective erythroid differentiation and maturation in Se deficiency [15]. Taken together, data suggest that the development of anemia caused by Se deficiency results from an excessive destruction of circulating erythrocytes along with a lack of sufficient production of mature erythroid cells.

Induction of inflammation

It has also been shown that oxidative stress mediated by increased hydrogen peroxide and lipid peroxides stimulates synthesis of IL-6 [39]. This proinflammatory cytokine has proven to be the most important direct inducer of hepcidin expression [40]. The Fe-regulatory hormone hepcidin, identified at the beginning of the XXIth century by Krause et al. [41], is a peptide predominantly produced by the hepatocytes [42, 43]. Its mechanism of action involves binding and inactivation of the transmembrane Fe exporter ferroportin [44].

Hepcidin plays a key role in the regulation of Fe homeostasis [45]. It suppresses the intestinal absorption of Fe, its release from macrophages recycling senescent erythrocytes and, at least partially, from hepatocytes [43]. Stimulated synthesis of hepcidin in inflammation thus results in a decreased Fe supply to erythropoiesis.

Elevated serum levels of IL-6 have been found in older women with low serum Se concentrations [46]. Se deficiency can therefore be considered to be an important factor in the pathogenesis of anemia of chronic inflammation mediated by the hormone hepcidin.

Increased expression of heme oxygenase-1

Deficiency of Se leads to increased oxidative stress and production of IL-6, both of which induce expression of heme oxygenase-1 [47, 48]. This enzyme catalyzes the initial reaction

of heme catabolism in which heme is reduced to biliverdin, carbon monoxide (CO), and free Fe²⁺.

Conclusions

Selenium deficiency has been associated with anemia in a number of studies involving animal and human subjects. At present, data of many studies support interactions between selenium and iron, although the exact underlying mechanisms by which selenium influences iron metabolism, synthesis of hemoglobin and erythropoiesis have not been fully established yet. One of the possible links between selenium and the homeostasis of iron is the peptide hepcidin. A more categorical clarification of the relationships between Se deficiency and development of anemia is needed with respect to appropriate trace element supplementation in cases of anemia with insufficient or absent therapeutic response to Fe treatment. Selenium supplementation could be considered in the strategies for treatment and control of anemia.

References

1. McLean E, Cogswell M, Egli I, Wojdyla D, Benoist B. Worldwide prevalence of anaemia, WHO Vitamin and Mineral Nutrition Information System, 1993–2005. *Public Health Nutr.* 2008;12(4):444-54. doi: 10.1017/S1368980008002401.
2. World Health Organization. The global prevalence of anaemia in 2011. Geneva: World Health Organization; 2015.
3. Van Nhien N, Khan NC, Yabutani T, Ninh NX, Chung LTK, Motonaka J, et al. Relationship of low serum selenium to anemia among primary school children living in rural Vietnam. *J Nutr Sci Vitaminol.* 2008;54:454-9.
4. Oldfield JE. Selenium world atlas (Updated edition). Grimbergen: Selenium-Tellurium Development Association (STDA); 2002.
5. Combs GF Jr. Selenium in global food systems. *Br J Nutr.* 2001;85(5):517-47.
6. Lozanov B, Tzachev K, Kirilov G, Aceva E, Vukov M. [Serum Selenium concentration and thyroid status in pregnant women on the background of systemic iodine supplementation]. *Endocrinologia.* 2008;13(2):56-64. Bulgarian.
7. Bivolarska A. [Iodine and iron deficiency in risk groups of population in some regions of Southern Bulgaria and influence of environmental factors on their distribution] [dissertation]. Plovdiv: Medical University – Plovdiv; 2014. Bulgarian.
8. Losanov B, Tzachev K, Kirilov G, Kovatcheva R, Aceva E, Apostolova E, Dobrikov P. [Selenium and thyroid status in children living in an endemic region with adequate iodine supplementation]. *Endocrinologia.* 2006;11(4):213-22. Bulgarian.
9. Moghadaszadeh B, Beggs AH. Selenoproteins and their impact on human health through diverse physiological pathways. *Physiology.* 2006;21:307-15. doi:10.1152/physiol.00021.2006.
10. Daniels LA. Selenium: does selenium status have health outcomes beyond overt deficiency? *Med J Aust.* 2004;180(8):373-4.
11. World Health Organization. Trace elements in human nutrition and health. Geneva: World Health Organization; 1996.
12. Rayman MP. Dietary selenium: time to act. *BMJ.* 1997;314(7078):387-8.
13. Chow C, Chen C. Dietary selenium and age-related susceptibility of rat erythrocytes to oxidative damage. *J Nutr.* 1980;110(12):2460-6.
14. Nagababu E, Chrest FJ, Rifkind JM. Hydrogen-peroxide induced heme degradation in red blood cells: the protective roles of catalase and glutathione peroxidase. *Biochim Biophys Acta.* 2003;1620:211-7.
15. Kaushal N, Hegde S, Lumadue J, Paulson RF, Prabhu KS. The regulation of erythropoiesis by selenium in mice. *Antioxid Redox Signal.* 2011;14(8):1403-12. doi: 10.1089/ars.2010.3323. PMID: PMC3061201.
16. Morris JG, Cripe WS, Chapman HL Jr, Walker DF, Armstrong JB, Alexander JD Jr, et al. Selenium deficiency in cattle associated with Heinz bodies and anemia. *Science.* 1984;223(4635):491-3.
17. Gutzwiller A. Erythrocyte resistance to oxidative damage and leucocyte capacity to reduce nitroblue tetrazolium in selenium-deficient cattle. *Zentralbl Veterinarmed A.* 1998;45(5):271-8.
18. Hu ML, Chung C, Spallholz JE. Hematologic data of selenium-deficient and selenium-supplemented rats. *J Inorg Biochem.* 1984;22(3):165-73.
19. Rashid N, Ghaznavi S. Association of levels of serum selenium with anemia in primary school children. *Biomedica.* 2015;31(2):128-31.
20. Van Nhien N, Yabutani T, Khan NC, Ninh NX, Motonaka J, Nakaya Y. Association of low serum selenium with anemia among adolescent girls living in rural Vietnam. *Nutrition.* 2009;25(1):6-10.
21. Bates CJ, Thane CW, Prentice A, Delves HT.

- Selenium status and its correlates in a British national diet and nutrition survey: people aged 65 years and over. *J Trace Elem Med Biol.* 2002;16(1):1-8.
22. Semba RD, Ferrucci L, Cappola AR, Ricks MO, Ray AL, Xue QL, et al. Low serum selenium is associated with anemia among older women living in the community: The Women's Health and Aging Studies I and II. *Biol Trace Elem Res.* 2006;112(2):97-107. doi:10.1385/BTER:112:2:97.
 23. Semba RD, Ricks MO, Ferrucci L, Xue QL, Guralnik JM, Fried LP. Low serum selenium is associated with anemia among older adults in the United States. *Eur J Clin Nutr.* 2009;63:93-9. doi:10.1038/sj.ejcn.1602889.
 24. Hampel G, Schaller K, Rosenmüller M, Oefele C. Selenium-deficiency as contributing factor to anemia and thrombocytopenia in dialysis patients. *Life Support Syst.* 1985;3(Suppl 1):36-40.
 25. VanLettow M, West C, vanderMeer J, Wieringa F, Semba R. Low plasma selenium concentrations, high plasma human immunodeficiency virus load and high interleukin-6 concentrations are risk factors associated with anemia in adults presenting with pulmonary tuberculosis in Zomba district, Malawi. *Eur J Clin Nutr.* 2005;59(4):526-32.
 26. Yetgin S, Hincal F, Başaran N, Ciliv G. Serum selenium status in children with iron deficiency anemia. *Acta Haematol.* 1992;88(4):185-8.
 27. Gürgöze MK, Aygün AD, Ölçücü A, Doğan Y, Yılmaz E. Plasma selenium status in children with iron deficiency anemia. *J Trace Elem Med Biol.* 2004;18(2):193-6. doi:10.1016/j.jtemb.2004.07.004.
 28. Gürgöze MK, Ölçücü A, Aygün AD, Taskin E, Kilic M. Serum and hair levels of zinc, selenium, iron, and copper in children with iron-deficiency anemia. *Biol Trace Elem Res.* 2006;111(1-3):23-9.
 29. Van Nhien N, Khan NC, Yabutani T, Ninh NX, Kasso A, Huong BTM, et al. Serum levels of trace elements and iron-deficiency anemia in adult Vietnamese. *Biol Trace Elem Res.* 2006;111(1-3):1-9.
 30. McAnulty LS, Gropper SS, McAnulty SR, Keith RE. Iron depletion without anemia is not associated with impaired selenium status in college-aged women. *Biol Trace Elem Res.* 2003;91(2):125-36.
 31. Viita LM, Mutanen ML, Mykkänen HM. Selenium-iron interaction in young women with low selenium status. *J Hum Nutr Diet.* 1989;2(1):39-42. doi: 10.1111/j.1365-277X.1989.tb00006.x.
 32. Lönnerdal B, Hernell O. Iron, zinc, copper and selenium status of breast-fed infants and infants fed trace element fortified milk-based infant formula. *Acta Paediatr.* 1994;83(4):367-73.
 33. Oster O, Schmiedel G, Prellwitz W. Correlations of blood selenium with hematological parameters in West German adults. *Biol Trace Elem Res.* 1988;15:47-81.
 34. Kaushal N, Hegde S, Gandhi U, Paulson RF, Prabhu KS. Selenium status and erythropoiesis in mice. *FASEB J.* 2009;23(1):728.5.
 35. Marinkovic D, Zhang X, Yalcin S, Luciano JP, Brugnara C, Huber T, et al. Foxo3 is required for the regulation of oxidative stress in erythropoiesis. *J Clin Invest.* 2007;117(8):2133-44.
 36. Padmaja K, Prasad AR. Selenium altered regulation of heme biosynthesis in chick embryos. *Drug Chem Toxicol.* 1993;16(4):395-408.
 37. Padmaja K, Ramamurthi R, Prasad AR. Inhibitory effect of selenium on enzymes involved in heme biosynthetic pathway in chick embryos. *J Enzyme Inhib.* 1996;11(1):1-11.
 38. Maciel EN, Bolzan RC, Braga AL, Rocha JB. Diphenyl diselenide and diphenyl ditelluride differentially affect delta-aminolevulinic acid dehydratase from liver, kidney, and brain of mice. *J Biochem Mol Toxicol.* 2000;14(6):310-9.
 39. Zhang J, Johnston G, Stebler B, Keller ET. Hydrogen peroxide activates NFkappaB and the interleukin-6 promoter through NFkappaB-inducing kinase. *Antioxid Redox Signal.* 2001;3(3):493-504.
 40. Nemeth E, Valore EV, Territo M, Schiller G, Lichtenstein A, Ganz T. Hepcidin, a putative mediator of anemia of inflammation, is a type II acute-phase protein. *Blood.* 2003;101(7):2461-3. doi:10.1182/blood-2002-10-3235.
 41. Krause A, Nietz S, Magert HJ, Schultz A, Forssmann WG, Schultz-Knappe P, et al. LEAP-1, a novel highly disulfide-bonded human peptide, exhibits antimicrobial activity. *FEBS Lett.* 2000;480:147-50.
 42. Kemna EH, Tjalsma H, Willems HL, Swinkels DW. Hepcidin: from discovery to differential diagnosis. *Haematologica.* 2008;93(1):90-7. doi:10.3324/haematol.11705.
 43. Nemeth E, Ganz T. The role of hepcidin in iron metabolism. *Acta Haematol.* 2009;122:78-86. doi: 10.1159/000243791.
 44. Nemeth E, Tuttle MS, Powelson J, Vaughn MB, Donovan A, Ward DM, et al. Hepcidin regulates cellular iron efflux by binding to ferroportin and inducing its internalization. *Science.* 2004;306(5704):2090-3.

45. Zhao N, Zhang AS, Enns CA. Iron regulation by hepcidin. *J Clin Invest.* 2013; 123(6):2337-43. doi: 10.1172/JCI67225. PubMed PMID: [23722909](#); PubMed Central PMCID: [PMC3668831](#).
46. Walston J, Xue Q, Semba R, Ferrucci L, Cappola A, Ricks M, et al. Serum antioxidants, inflammation, and total mortality in older women. *Am J Epidemiol.* 2006;163(1):18-26.
47. Tron K, Novosyadlyy R, Dudas J, Samoylenko A, Kietzmann T, Ramadori G. Upregulation of heme oxygenase-1 gene by turpentine oil-induced localized inflammation: involvement of interleukin-6. *Lab Invest.* 2005;85(3):376-87.
48. Mostert V, Nakayama A, Austin LM, Levander XA, Ferris CD, Hill KE, et al. Serum iron increases with acute induction of hepatic heme oxygenase-1 in mice. *Drug Metab Rev.* 2007;39(2-3):619-26.